

ehp

Environmental Health

ehponline.org

P E R S P E C T I V E S

Published by the National Institute of
Environmental Health Sciences

Genetic Factors that Might Lead to Different Responses in Individuals Exposed to Perchlorate

**Franco Scinicariello, H. Edward Murray,
Lester Smith, Sharon Wilbur, and Bruce A. Fowler**
doi:10.1289/ehp.8076 (available at <http://dx.doi.org/>)
Online 29 June 2005



**The National Institute of Environmental Health Sciences
National Institutes of Health
U.S. Department of Health and Human Services**

**Genetic Factors that Might Lead to Different Responses
in Individuals Exposed to Perchlorate**

**Franco Scinicariello, H. Edward Murray, Lester Smith,
Sharon Wilbur, Bruce A. Fowler.**

Division of Toxicology, Agency for Toxic Substances and Disease Registry, Centers for
Disease Control and Prevention. Atlanta, Georgia 30333, USA

Correspondence to:

Franco Scinicariello, MD, MPH

Division of Toxicology, ATSDR, CDC

4770 Buford Highway, MS: F-32,

Atlanta, Georgia 30341, USA

Phone: 770-488-3331

Fax: (770)-488-4178

E-mail: fes6@cdc.gov

Running Title: Genetic Factors and Perchlorate.

Key words: Perchlorate, Genetic Susceptibility, Mutations, Thyroid Gland, Hypothyroidism, NIS, Pendred Syndrome, Pendrin, TPO

Acknowledgements

We thank O. Harris for critical comments.

This project was supported under a cooperative agreement from the Centers for Disease Control and Prevention through the Association of Teachers of Preventive Medicine.

Franco Scinicariello is a recipient of an ATPM Career Development Award.

The authors declare they have no competing financial interests.

Abbreviations:

CH = Congenital hypothyroidism;
DIT = Diiodo-tyrosines;
ITD = Iodide transport deficit;
MIT = Monoiodo-tyrosines;
NIS = Sodium-iodide symporter;
PIOD = Partial iodide organification defects;
PDS = Pendred syndrome;
RAIU= Radioactive iodine uptake;
T₃ = Triiodothyronine;
T₄ = Thyroxine;
Tg = Thyroglobulin;
TH = Thyroid hormone;
TIOD = Total iodide organification defects;
TPO = Thyroid peroxidase;
TRH = Thyrotropin-releasing hormone;
TSH = Thyroid-stimulating hormone;
TSHr = TSH receptor.

Abstract

Introduction

Public health and perchlorate

Mode of action of perchlorate in human

Thyroid hormone synthesis

Defects in iodide transport circulation into the thyroid cell

**Defects in iodide transport from the thyroid cell to the follicular lumen, often
combined with inner ear deafness (Pendred syndrome)**

Defects of iodide organification

Relevant studies of perchlorate in humans

Conclusions

Abstract

Perchlorate has been detected in groundwater in many parts of the United States and recent detection in vegetable and dairy food products indicates that contamination by perchlorate is more widespread than previously thought. Perchlorate is a competitive inhibitor of the sodium-iodide symporter, the thyroid cell surface protein responsible for transporting iodide from the plasma into the thyroid. An estimated 4.3 % of the U.S. population is subclinically hypothyroid, and 6.9% of pregnant women may have low iodine intake. Congenital hypothyroidism affects 1:3000–1:4000 infants, and 15% of these cases have been attributed to genetic defects. The objective of this review is to identify genetic biomarkers that would help to define sensitive subpopulations to environmental perchlorate exposure. We reviewed the literature to identify genetic defects involved in the iodination process of the thyroid hormone synthesis particularly: defects in iodide transport from circulation into the thyroid cell; defects in iodide transport from the thyroid cell to the follicular lumen (Pendred syndrome); and defects of iodide organification. Furthermore, we summarized relevant studies of perchlorate in humans. Because of perchlorate inhibition of iodide uptake, it is biologically plausible that chronic ingestion of perchlorate through contaminated sources may cause some degree of iodine discharge in populations that are genetically susceptible to defects in the iodination process of the thyroid hormone synthesis, thus deteriorating their conditions. We conclude that future studies linking human disease and environmental perchlorate exposure should consider the genetic makeup of the participants, actual perchlorate exposure levels, and individual iodine intake/excretion levels.

Introduction

Sequencing of the human genome has brought new emphasis and increased interest in gene-environment interactions and is becoming relevant in defining public health policies. For many years, people's susceptibility to xenobiotics has been known to differ significantly. Now, several techniques are available to identify and characterize the genetic correlates of inter-individual variability. The goal of environmental genomics is to understand how genetic variability influences individual responses to environmental factors on the basis of assumption that high-risk genotypes accumulate more damage and, therefore, are at greater risk of developing exposure-related diseases. Thus, genomics information may lead to development of predictive biomarkers that identify potentially sensitive populations and earlier prediction of adverse outcomes, ultimately resulting in better intervention strategies (Kelada 2003).

Public health and perchlorate

The advent and use of new, highly sensitive detection techniques has identified contamination of groundwater by perchlorate in many parts of the United States, primarily in association with industries involved in rocket, explosives, and fireworks manufacturing and propellant handling. Concentrations measured in most public water supplies are $<50 \mu\text{g/L}$; however, levels as high as several hundred $\mu\text{g/L}$ have been reported in some drinking water wells in certain communities (Motzer 2001). The recent detection of perchlorate in vegetable and dairy food products (Kirk et al. 2003; Smith et al. 2001) indicates that contamination by perchlorate is more widespread than previously thought.

Perchlorate (ClO_4^-) is the dissociated anion of perchlorate salts such as potassium perchlorate, sodium perchlorate, and ammonium perchlorate and is extremely water-soluble and environmentally stable. Therefore, the perchlorate ion is identical whether derived from potassium, sodium, or ammonium salts. Potassium perchlorate was used primarily as a pharmaceutical agent to treat hyperthyroidism. It is now used mainly in flares and automobile airbags, although it is still used for diagnostic purposes and for treatment of hyperthyroidism. Sodium perchlorate is used in the manufacture of slurry explosives. Ammonium perchlorate is widely used as a component of propellants for rockets, missiles, and fireworks (Soldin et al. 2001)

Perchlorate was first detected in high concentrations by monitoring wells in California during the early 1990s. When initially detected in California, the Region 9 Office of the U.S. Environmental Protection Agency developed a provisional reference dose for perchlorate of 0.0001 mg/kg-day in 1992. The reference dose later was revised to 0.0007 mg/kg-day in 2005. These values were based on a dated acute exposure study, which showed that single doses of potassium perchlorate caused release of iodide (I^-) from thyroids of patients with Graves disease, an autoimmune condition that results in hyperthyroidism (Stanbury and Wyngaarden 1952).

Mode of action of perchlorate in human

Perchlorate is a competitive inhibitor of the sodium-iodide symporter (NIS), the thyroid cell surface protein responsible for transporting iodide from the plasma into the thyroid. Therefore, it prevents further synthesis of the thyroid hormone (TH). It has no effect on the iodination process itself; rather, it displaces iodide by competitive uptake at

the NIS. Perchlorate is concentrated by the thyroid tissue in a manner similar to iodide, but it is neither significantly metabolized in the gland nor peripherally (Wolff 1998). Eskandary et al (1997) disputed the notion that perchlorate is translocated via NIS into the cell, and that perchlorate acts on NIS as a blocker, not as a substrate. Therefore, it is possible that perchlorate may cross the thyrocyte membrane by diffusion. In rats and humans, perchlorate appears to be eliminated rapidly, primarily in urine (>90%), virtually unchanged (Anbar et al. 1959; Eichler and Hackenthal 1962).

Several other inorganic anions, which are present in dietary and environmental sources, such as thiocyanate (SCN^-) and nitrate (NO_3^-) have goitrogenic effects (Greer et al. 1966). Similarly to perchlorate, they both competitively inhibit iodide uptake at NIS. Several studies have been conducted to determine the relative effects of perchlorate, thiocyanate and nitrate on radioactive iodide uptake (RAIU) inhibition. Studies in rats showed that perchlorate was approximately 10 times more potent than thiocyanate, and about 300 times more potent than nitrate in inhibiting RAIU in the thyroid. Furthermore, thiocyanate was slightly more potent than iodide (Wyngaarden et al. 1953). Tonacchera et al (2004) demonstrated, in Chinese hamster ovary (CHO) cell lines stably transfected with human *NIS*, that the relative potency of perchlorate on RAIU inhibition was 15, 30 and 240 times that of thiocyanate, iodide and nitrate, respectively. The inhibiting effects when the cell lines were exposed to a mixture of perchlorate, thiocyanate and nitrate were simply additive.

Thyroid hormone synthesis

TH plays a key role in the growth and differentiation of many organs. It is especially important for development of the central nervous system during the prenatal and postnatal periods (reviewed in Zoeller et al. 2002). A severe shortage of TH for several weeks after birth results in serious mental and motor handicaps. During pregnancy, the mother provides substantial amounts of TH to the fetus (Vulsma et al. 1989), so the delay in cerebral development caused by congenital hypothyroidism (CH) results mainly from postnatal TH deficiency. The risk for mental retardation and the difficulty in recognizing the disease were reasons for introducing neonatal mass screening programs. Therefore, the most serious effects of perchlorate might occur during the first trimester when the brain is forming and developing and thyroid hormone supply is totally dependent on maternal supply of iodine and of thyroxine (T_4) and triiodothyronine (T_3)

To understand the potential impact of perchlorate on a gene-environment interaction model, we need to consider the THs, T_3 and T_4 , in a proper biosynthesis context. TH synthesis and secretion are exquisitely regulated negative-feedback systems that involve the hypothalamus, pituitary, and thyroid glands. The hypothalamus secretes thyrotropin-releasing hormone (TRH), a tripeptide (PyroGlu-His-Pro) synthesized in the paraventricular nucleus of the hypothalamus. The TRH, transported by axons, binds to TRH receptors in the pituitary thyrotropes, a subpopulation of pituitary cells that secrete thyroid-stimulating hormone (TSH). TRH stimulation leads to release and synthesis of new TSH in thyrotropes. The TSH binds to the TSH receptor (TSHr) in the thyroid gland cells. TSH is the primary regulator of TH release and secretion. Both TRH and TSH

secretion are negatively regulated by THs: when T_4 reaches an adequate circulating level, the hypothalamus and pituitary reduce their output of TRH and TSH and increase their output of TRH and TSH when the circulating blood level of T_4 is low. A number of thyroid genes, including *NIS*, thyroglobulin (*Tg*), and thyroid peroxidase (*TPO*), are stimulated by TSH and promote the synthesis of TH (Zoeller 2003).

Iodine is critical to thyroid gland function and TH synthesis and secretion. The first step in thyroidal iodine metabolism is the cellular uptake of iodide from the extracellular fluid. The thyroidal iodine uptake is tightly regulated by the NIS, an intrinsic plasma membrane protein in the thyroid follicular cells (Dohán et al. 2003). From the follicular cell, the iodide moves across the apical membrane transported by pendrin protein (Yoshida et al. 2002). The iodide is then delivered to the cell-colloid interface where it is oxidized by TPO and bound to tyrosyl residues in the Tg. This iodination of specific tyrosines on Tg yields monoiodinated and diiodinated residues (MIT, monoiodo-tyrosines; DIT, diiodo-tyrosines) that are enzymatically coupled to form T_4 and T_3 . The iodinated Tg containing MIT, DIT, T_4 , and T_3 , then is stored as an extracellular storage polypeptide in the colloid within the lumen of thyroid follicular cells.

Perchlorate does not undergo metabolism, but genetic defects of its target, i.e., the NIS, may lead to low iodine uptake in the thyroid gland, thus depressing production of THs. In this scenario, exposure to perchlorate may further reduce the already low iodide uptake and decrease production of THs. The combined effects of perchlorate with a genetic decrease in THs would hence delineate a population at risk for decreased thyroid function.

We reviewed published data to identify genetic factors that might lead to different responses in people exposed to perchlorate in the environment. Because perchlorate inhibits iodide uptake, we focused on the genetic defects causing congenital hypothyroidism involving the iodination process of the THs, particularly: i) defects in iodide transport from circulation into the thyroid cell; ii) defects in iodide transport from the thyroid cell to the follicular lumen, often combined with inner ear deafness (Pendred syndrome [PDS]); and; iii) defects of iodide organification.

A positive perchlorate discharge test is used as diagnostic tool in most of these medical conditions. A positive diagnosis can be obtained by administering 1 gm potassium perchlorate two hours after a tracer dose of I^{131} . In normal individuals, radioiodide accumulation in the thyroid gland comes to an end after the administration of potassium perchlorate, but there is little loss of the thyroidal radioactivity previously accumulated in the gland. Instead, potassium perchlorate causes almost complete discharge of the unbound fraction of thyroid iodide in individuals with defects of iodide organification and with PDS. Therefore, these people could have a different response to environmental perchlorate exposure than normal individuals.

Defects in iodide transport from circulation into the thyroid cell

The NIS is the plasma membrane glycoprotein that mediates active iodide uptake into the thyroid follicular cells. This process is the crucial first step in TH biosynthesis. NIS couples the inward transport of sodium, which occurs in favor of its electrochemical gradient, to the simultaneous inward translocation of iodide against its electrochemical gradient. Two sodium ions per iodide ion are translocated into the cells (Dai et al. 1996;

Eskandari et al. 1997). The sodium gradient that drives iodide uptake is maintained by the Na⁺/K⁺ ATPase.

Congenital iodide transport deficit (ITD) is an infrequent autosomic recessive condition characterized by inability of the thyroid gland to maintain a concentration gradient of iodide between the plasma and the thyroid follicular cell, resulting in hypothyroidism, diffuse or nodular goiter, and little or no uptake of radioiodine. The disorder has been linked to a defect of the NIS. In the absence of a functional NIS molecule, iodide has no access to the thyroid follicular cells, resulting in decreased TH biosynthesis and higher circulating levels of TSH, which in turn stimulates the morphologic and biochemical changes in the thyroid that result in development of goiter (De La Vieja et al. 2000).

The gene coding for human NIS has been mapped to chromosome 9p12-13.2. It has 15 exons and coding for a glycoprotein of 643 amino acids. NIS is a protein with 13 putative transmembrane domains, an extracellular amino terminus and an intracellular carboxylterminus (De La Vieja et al. 2000). About 58 cases of ITD from 33 families have been reported worldwide. Thirty of 31 cases from 21 families were studied at the molecular level and had several homozygous or compound heterozygous mutations of the perchlorate-sensitive *NIS* gene. Eleven mutations have been identified, namely V59E, G93R, Q267E, C272X, T354P, G395R, frame-shift 515X, Y531X, G543E, ΔM142-Q323, and ΔA439-P443 (Fujiwara et al. 1997, 1998, 2000; Kosugi et al. 1998a, 1998b, 1999, 2002; Matsuda et al. 1997; Pohlenz et al. 1997, 1998; Tonacchera et al. 2003). The single substitution in the 354th codon converting from ACA (Thr) to CCA (Pro) was the most common mutation detected in 10 patients with homozygous mutations, and in four

patients with compound heterozygous mutation (Fujiwara et al. 1997, 1998; Matsuda et al. 1997; Kosugi et al. 1998a, 1998b). All were Japanese, suggesting that the mutant NIS T354P is more common in Japan. However, the frequency of this gene in the Japanese population is unknown because only 185 healthy people, representing only 370 alleles, have been genotyped.

The frequency of mutations in the *NIS* gene in the population is not known. Heterozygous persons do not express the phenotype; therefore, *NIS* gene defects can be detected only when both alleles are affected. People with homozygous mutations that cause partial loss of function may not be detected when, under conditions of high iodide intake, full preservation of iodide concentrating function is not required to achieve normal hormone synthesis. Therefore, impairment of thyroidal iodide concentration requires not only mutations in both *NIS* alleles but also defects that cause virtually complete loss of function.

The therapeutic treatment of ITD patients consists of L-T₄ administration. Some patients also are supplemented with potassium iodide, thus underscoring the degree of functional loss of the mutated NIS. In these persons, perchlorate intake from contaminated sources could further reduce the functional activity of the mutated NIS in concentrating iodide in the thyroid.

Defects in iodide transport from the thyroid cell to the follicular lumen, often combined with inner ear deafness (Pendred syndrome).

PDS, an autosomal recessive disorder characterized by deafness and goiter, is the most common cause of syndromic deafness, accounting for up to 10% of all hereditary

hearing loss (Fraser 1965; Nilsson et al. 1964). A phenotypic heterogeneity exists among affected persons, and thyroid dysfunction is particularly variable. Whereas at least 50% of affected persons have normal circulating levels of TH, others develop clinical hypothyroidism (Reardon et al. 1999). Most affected persons demonstrate impaired iodide organification, as determined by a positive perchlorate discharge test. Hearing loss in PDS is prelingual, and in at least 80% of patients, is associated with structural defects of the inner ear, including a dilatation of the vestibular aqueduct and the Mondini defect of the cochlea (Johnsen et al. 1989; Phelps et al. 1998). The PDS gene (*SLC26A4*), has been linked to chromosomal region 7q31 and contains an open reading frame of 2343bp encompassing 21 exons (Coyle et al. 1996; Sheffield et al. 1996) The predicted gene product, pendrin, is a highly hydrophobic 780 amino acid protein that transports chloride and iodide and mediates the exchange of chloride and formate. In the thyroid gland, a disorder in the function of pendrin may cause diminished iodide transport over the apical membrane that results in iodide remaining in the thyrocyte and a consequent decrease of organification of iodide. As a result, iodide accumulates in the cytoplasm and is discharged if thiocyanate or perchlorate is given (perchlorate discharge test). A decrease in the amount of radiolabeled iodide over the thyroid of >10% is considered positive. At least 85 independent *SLC26A4* gene mutations have been characterized as causing PDS and nonsyndromic deafness, in some cases confirmed by a normal perchlorate discharge test (Adato et al. 2000; Blons et al. 2004; Bogazzi et al. 2000, 2004; Campbell et al. 2001; Coucke et al. 1999; Coyle et al. 1998; Everett et al. 1997; Fugazzola et al. 2000; Kopp et al. 1999; Li et al. 1998; Lopez-Bigas et al. 2002; Namba et al. 2001; Park et al. 2003; Phelps et al. 1998; Prasad et al. 2004; Reardon et al. 2000; Scott et al. 2000; Tekin

et al. 2003; Tsukamoto et al. 2003; Usami et al. 1999; Van Hauwe et al. 1998; Yong et al. 2001). Although these mutations are distributed throughout the coding sequence, having been identified in 19 of the 21 exons, the spectrum of mutations appears to show geographic differences. In Caucasian patients, the L236P, T416P, and IVS8+1G>A mutations account for nearly half of all *SLC26A4* mutant alleles, whereas in Japanese patients, these mutations are rare (Campbell et al. 2001; Tsukamoto et al. 2003). By contrast, H723R and ISV7-2A>G are the prevalent alleles accounting for most observed *SLC26A4* mutations in Korean and Japanese studies (Park et al. 2003; Tsukamoto et al. 2003). Some researchers have suggested that the frequency of these mutations could represent a founder effect, rather than mutational hot spots.

A disorder in the function of the pendrin will cause a diminished iodide transport over the apical membrane, which causes iodide to remain in the thyrocyte. Intake of perchlorate from contaminated source may cause discharge of iodide from the thyrocyte further exacerbating the organification defect with resulting decrease of TH synthesis.

Moreover, at the present, it is not known whether perchlorate will affect the function of the normal pendrin protein to transport iodide. Molecular studies addressing whether perchlorate may act on iodide transport through inhibition of the pendrin protein in a fashion similar to the NIS are needed and welcomed.

Defect in iodide organification

Iodide organification is the process by which iodine is oxidized and bound to thyrosine residue in Tg. Thyroid iodide organification disorder represents a group of defects characterized by discharge of substantial percentage of labeled iodide from the

thyroid after administration of perchlorate (perchlorate discharge test) or thiocyanate. This discharge indicates a defect in converting accumulated iodide to organically bound iodine. The discharge may be partial or complete, thus defining partial or total defects. Partial iodide organification defects (PIODs) are characterized by release of <50% of the accumulated radioiodine. Total iodide organification defects (TIODs) are characterized by release of >90% of the accumulated radioiodine.

Iodination of the tyrosine residue is catalyzed by the membrane-bound thyroperoxidase (TPO). However, the oxidation of iodine requires hydrogen peroxide synthesized outside the thyroid follicular cell at the apical border catalyzed by the thyroid complex. Recently, two proteins of this complex DUOX1 (also known as THOX1) and DUOX2 (also known as THOX2) have been identified (De Deken et al. 2000; Dupuy et al. 1999.). The *DUOX1* and *DUOX2* genes are co-localized on 15q15.3 chromosome and code for proteins of 1551 and 1548 amino acids, respectively. The DUOX1 and DUOX2 structure includes seven transmembrane-spanning domains, three NADPH- and one FAD-binding site, and 2EF-hand motifs. During the past 3 decades, few cases of thyroidal hydrogen peroxide have been described, but the molecular bases of these defects have just recently been investigated. Moreno et al. (2002) reported mutations in the *DUOX2* gene, resulting in premature stop codon, in four CH patients with unexplained iodide organification defects. One patient with permanent CH and TIOD carried a homozygous substitution, whereas three patients with temporary CH and PIOD carried heterozygous mutations that cause premature termination signal.

Lack of or insufficient activity of the DUOX2 protein diminishes hydrogen peroxide production, resulting in decreased activity of TPO and accumulation of iodide in

the thyrocyte. Intake of environmental perchlorate, which inhibits iodine inflow, also may cause discharge of unbound iodine, further deteriorating the iodine organification process.

Under oxidative conditions, TPO catalyzes the coupling of iodotyrosines to iodothyronine residue in Tg. Thyroperoxidase is a glycosylated hemoprotein encoded by the *TPO* gene located on chromosome 2p25. The gene contains 17 exons coding for a protein of 933 amino acids. The protein has a transmembrane helix with a large extracellular N-terminal part containing a heme group. TPO defects are believed to be among the most frequent causes of abnormalities in thyroid iodide organification defect causing goitrous congenital hypothyroidism. TPO activity is not detectable in thyroid tissue of patients with TIOD. Absence of TPO activity implicates the inability to iodinate tyrosine residue in Tg and to couple these residues to form THs, mainly T₄ and some T₃ and r T₃. Inactivating mutations in both *TPO* alleles have been found in patients with congenital hypothyroidism caused by TIOD. With use of a variety of molecular techniques for mutation detection, 36 mutations have now been defined for *TPO*. These include frameshift mutations caused by nucleotide insertion or deletion, as well as missense, nonsense, and splice site mutations (Abramowicz et al. 1992; Ambrugger et al. 2001; Bakker et al. 2000; Bikker et al. 1994, 1995, 1997; Kotani et al. 2001; Nascimento et al. 2002; Niu et al. 2002; Pannain et al. 1999; Rivolta et al. 2003; Santos et al. 1999; Umeki et al. 2002, 2004; Wu et al. 2002). The first reported mutation was a homozygous GGCC insertion in exon 8 of the *TPO* gene. The resulting frameshift generates a stop codon in exon 9, which results in a grossly truncated protein with no expected activity (Abramowicz et al. 1992). In a Dutch study of 45 patients from 40 families with CH

caused by TIOD, the GGCC insertion in exon 8 at nucleotide position 1287 was the most common mutation found (Bakker et al. 2000). It was detected in 36% of the investigated *TPO* alleles and in 51% of the families investigated either in a homozygous or a compound heterozygous fashion. In this study, mutations in both *TPO* alleles were found in 29 families: for 13 families in a homozygous fashion and for 16 families in a compound heterozygous fashion. In total, 16 different mutations were found, including eight novel mutations: six frameshift mutations, six missense mutations, three splice site mutations, and one nonsense mutation. Most of these mutations occurred in exon 8, 9, or 10, which encode for the active part of the enzyme involved in the heme binding. In one patient with classic TIOD, a homozygous deletion in exon 14 appeared to have resulted from partial maternal isodisomy of the short arm of chromosome 2 carrying the defective *TPO* gene (Bakker et al. 2001). In some patients, alternative splicing would generate a partially active form of the enzyme. In others, an early termination signal would prevent translation of the fully active protein (Abramovicz et al. 1992; Bikker et al. 1994, 1995; Manglabruks et al. 2001; Santos et al. 1999). Umeki et al. (2002) described two novel mutations in the *TPO* gene, R665W and G771R, in exons 11 and 13, respectively. The former was found in the patient's father (heterozygous) and the latter in her mother, also heterozygous. No TPO activity was detectable with cells transfected with mutated mRNAs. Moreover, the mutated TPO proteins showed abnormal cellular localization, exhibiting immunofluorescence only in the intracellular structure. Therefore, the loss of apical membrane localization of the mutated TPO was the main cause for the iodide organification defect.

PIODs also can be caused by disorders in TPO. In an investigation of *TPO* mutations in five families with PIOD, Nascimento et al. (2002) found a compound heterozygous mutation in three patients from one family inherited from both heterozygous parents. In the other four families, they found only heterozygous *TPO* mutations or polymorphisms, suggesting the translated protein could be partially inactive. Recently, PIOD caused by *TPO* gene was diagnosed in three siblings (Kotani et al. 2003). The three siblings with goiter and latent to mild hypothyroidism had a compound heterozygous mutation for a missense mutation (G1687T) and a deletion in exon 10 (1808–13del) resulting in a produced protein with two deleted amino acids Δ D574-L4575. From the expression studies, the mutated Δ D574-L4575 -TPO, synthesized THs to some extent (Kotani et al. 2003).

A common feature of patients with thyroid organification disorders syndrome is the discharge of iodine from the thyroid after administration of perchlorate. The level of perchlorate administered in the diagnostic test is higher than the reported level of contaminated sources. However, it is biologically plausible that cumulative ingestion of perchlorate through contaminated source, may cause some degree of iodine discharge from thyrocytes. In populations with partial activity of the TPO enzyme, exposure to high enough environmental perchlorate could cause unbound iodide discharge; therefore, less iodine will be available for biosynthesis of THs, thus further deteriorating their conditions.

Relevant studies of perchlorate in humans

Many studies have attempted to provide useful information on the dose-response relation of perchlorate-related health effects. Several ecologic studies have compared thyroid function in newborns using T₄ and TSH screening data in infants born to mothers in areas with different perchlorate exposure. However, these studies yielded contradictory results. Brechner et al. (2000) found higher TSH in newborns in Yuma, Arizona, which has high perchlorate exposure, than in Flagstaff, Arizona, which has lower exposure. However, whether perchlorate exposure caused the observed TSH effect cannot be addressed because of the lack of direct perchlorate measurement in the study. By contrast, Li et al. (2000a, 2000b) found no association in Nevada newborns between low T₄ and TSH levels and perchlorate exposure. A limitation of these studies is that the investigators did not collect data on individual exposure to perchlorate and on iodine intake levels. In an unpublished population-based ecologic study using California Newborn Screening Program data, Schwartz (2001) claimed to identify a significant dose-response association between perchlorate exposure and T₄, and an association of perchlorate exposure and being a presumptive positive for congenital hypothyroidism. These data contrast with a previous ecologic analysis (Lamm and Doemland 1999), that found no increase of congenital hypothyroidism incidence in California and Nevada counties with perchlorate levels of 4–16 µg/L in drinking water supplies.

Crump et al. (2000) conducted a study in three proximate cities in northern Chile, which had different concentrations of perchlorate in tap water, involving 162 school-aged children and 9,784 newborns. These authors found no alteration of thyroid function or incidence of congenital hypothyroidism in Taltal, Chile, where the tap water contained

100–120 µg/L perchlorate compared with two other regions of Chile with low or no perchlorate in the water. However, the data also showed high levels of urine iodine, indicating that iodine intake in the population was very high, possibly overcoming the inhibitory effect of perchlorate on thyroid function.

To establish the dose response in humans for the perchlorate inhibition of thyroidal iodide uptake and the short-term effects on circulating TH, Greer et al. (2002) gave perchlorate in drinking water at 0.007, 0.02, 0.1, or 0.5 mg/kg per day to 37 male and female volunteers for 14 days. In 24 participants, 8- and 24-hr measurements of thyroidal ¹²³I uptake (RAIU) were performed before exposure, on exposure days 2 and 14, and 15 days postexposure. Results from the study indicated a true no-effect level of perchlorate of 5.2 or 6.4 µg/kg/day for RAIU. Considering that a 70 kg adult drinks 2 liters of water per day, this dose would be ingested if the drinking water contained 182–224 µg/L. In addition, the dose of 0.5 mg/kg/day taken for 14 days did not produce changes in circulating levels of T₄ or TSH, suggesting that short term consumption of perchlorate levels of 17.5 mg/L in drinking water would not affect circulating levels of THs. The authors suggested that this failure of perchlorate to influence circulating levels of TH resulted from the storage capacity of the normal adult thyroid gland, which contains unreleased stored hormones lasting for several months. However, as pointed out by Zoeller (2003), the case may be different for a late gestation fetus or neonate, where the estimated intra-thyroidal amount of hormone stored is less than that required for 1 day (Van den Hove et al. 1999; Vulsma et al. 1989). Thus, the concentration of perchlorate sufficient to reduce thyroidal iodine uptake in a fetus or neonate may be sufficient to produce a significant decrement in circulating levels of TH. The fetal thyroid gland

obtains iodide for its own TH synthesis from the maternal circulation through the placenta. Placental transfer of perchlorate has been reported in guinea pig (Postel 1957). In human, whether perchlorate crosses from the mother to the fetus during pregnancy is not known. However, this placental transfer could be biologically plausible because expression of the *NIS* has been reported in human placenta (Bidart et al. 2000). Moreover, perchlorate may concentrate in milk because the NIS protein is induced in lactating breast tissue by prolactin (Tazebay et al. 2000). Perchlorate might decrease iodide uptake into milk, thus reducing the sole source of iodine to the infant. Differently from adults, who most likely can recover from transient hypothyroidism without permanent health consequences, a short period of TH insufficiency may produce permanent neurologic deficits in children (Van Vliet 1999). The study of no-effect level (Greer et al. 2002) was conducted in healthy adults with normal iodine intake and it is debatable whether 14 days is sufficient time to illustrate perchlorate effect on humans. This no-effect level most likely would be lower in populations with genetic defects causing CH and in populations with lower iodine uptake. The third National Health and Nutrition Examination Survey (NHANES III), conducted during 1988–1994, found that the percentages of males and females with urinary iodine (UI) concentrations $<5 \mu\text{g/dL}$ were substantially higher in every age category than in the 1971–1974 survey (Hollowell et al. 2002). In pregnant women, these percentages were 6.9% in NHANES III and 1.0% in NHANES I. The overall decline in the last few decades raises concern that a fairly large number of people in the United States may lack adequate iodine intake.

Conclusions

Exposure to perchlorate, which inhibits iodine uptake, has the biological potential to cause hypothyroidism and, in pregnant women, severely damage the fetus and the newborn. NHANES III data suggest that 4.3% of the U.S. population may be subclinically hypothyroid (Hollowell et al. 2002). Congenital hypothyroidism affects about 1:3000–1:4000 infants and in about 15% of cases may result from a defect of thyroid hormonogenesis, mostly inherited in an autosomal recessive fashion (Vulsma and de Vijlder 2000). Such defects may result from abnormalities in several steps involved in TH synthesis. Our literature review identified possible homozygous or compound heterozygous mutations of genes involved in thyroid iodine synthesis that cause hypothyroidism that could be used to define a potential susceptible population to perchlorate exposure. In a Mendelian fashion, the number of carriers of heterozygous mutated gene causing CH would be higher than the number of the reported CH cases. Given the logical connection between perchlorate, diminished iodine uptake, hypothyroidism, and thyroid-related health effects, people exhibiting heterozygous or homozygous genetic mutations in genes involved in the TH synthesis, especially in a milieu of low iodine uptake, can reasonably be expected to be more susceptible than people who show no genetic variability to the effects of perchlorate. Several studies based on T₄ and TSH screening data in infants born to mothers in areas with different perchlorate exposure mostly have found no increase in hypothyroidism incidence. However, these studies lacked estimates of individual perchlorate exposure, as well as estimates of individual iodine uptake. The only study that included iodine values showed no significant association between perchlorate and hypothyroidism. However, it showed

high urinary iodide, suggesting the high iodine uptake could easily have upset the inhibition factor of the perchlorate. We conclude that future epidemiologic and population-based studies, as well as no-effect studies, concerning link between human disease and environmental perchlorate exposure should consider among their variables the genetic makeup of the participants, actual perchlorate exposure levels, and individual iodine uptake and excretion levels.

References

Abramowicz MJ, Targovnik H, Varela V, Cochaud P, Krawiec L, Pisarev MA, et al. 1992. Identification of a mutation in the coding sequence of the human thyroid peroxidase gene causing congenital goiter. *J Clin Invest* 90:1200–1204.

Adato A, Raskin L, Petit C, Bonne-Tamir B. 2000. Deafness heterogeneity in a Druze isolate from the Middle East: novel OTOF and PDS mutations, low prevalence of GJB2 35delG mutation and indication for a new DFNB locus. *Eur J Hum Genet* 8:437–442.

Ambrugger P, Stoeva I, Biebermann H, Torresani T, Leitner C, Gruters A. 2001. Novel mutations of the thyroid peroxidase gene in patients with permanent congenital hypothyroidism. *Eur J Endocrinol* 145:19–24.

Anbar M, Guttman S, Lweitus Z. 1959. The mode of action of perchlorate ions on the iodine uptake of the thyroid gland. *Int J Appl Radiat Isot* 7:87–96.

Bakker B, Bikker H, Vulsma T, de Randamie JS, Wiedijk BM, de Vijlder JJM. 2000. Two decades of screening for congenital hypothyroidism in The Netherlands: *TPO* gene mutations in total iodide organification defects (an update). *J Clin Endocrinol Metab* 85:3708–3712.

Bakker B, Bikker H, Hennekam RC, Lommen EJ, Schipper MG, Vulsma T, et al. 2001. Maternal isodisomy for chromosome 2p causing severe congenital hypothyroidism.

J Clin Endocrinol Metab 86:1164–1168.

Bidart JM, Lacroix L, Evain-Brion D, Caillou B, Lazar V, Frydman R, et al. 2000. Expression of Na⁺/I⁻ symporter and Pendred syndrome genes in trophoblast cells. J Clin Endocrinol Metab 85:4367–4372.

Bikker H, Den Hartog MT, Baas F, Gons MH, Vulsma T, de Vijlder JJM. 1994. A 20 base pair duplication in the human thyroid peroxidase gene results in a total iodide organification defect and congenital hypothyroidism. J Clin Endocrinol Metab 79:248–252.

Bikker H, Vulsma T, Baas F, de Vijlder JJM. 1995. Identification of five novel inactivating mutation in the human thyroid peroxidase gene by denaturing gradient gel electrophoresis. Hum Mutat 6:9–16.

Bikker H, Baas F, de Vijlder JJM. 1997. Molecular analysis of mutated thyroid peroxidase detected in patients with total iodide organification defects. J Clin Endocrinol Metab 82:649–653.

Blons H, Feldmann D, Duval V, Messaz O, Denoyelle F, Loundon N, et al. 2004. Screening of *SLC26A4* (PDS) gene in Pendred's syndrome: a large spectrum of mutations in France and phenotypic heterogeneity. Clin Genet 66:333–340.

Bogazzi F, Raggi F, Ultimieri F, Campomori A, Cosci C, Berrettini S, et al. 2000. A novel mutation in the pendrin gene associated with Pendred's syndrome. *Clin Endocrinol* 52:279–285.

Bogazzi F, Russo D, Raggi F, Ultimieri F, Berrettini S, Forli F, et al. 2004. Mutations in the *SLC26A4* (pendrin) gene in patients with sensorineural deafness and enlarged vestibular aqueduct. *J Endocrinol Invest* 27:430–435.

Brechner R, Parkhurst G, Humble W, Brown M, Herman W. 2000. Ammonium perchlorate contamination of Colorado River drinking water is associated with abnormal thyroid function in newborns in Arizona. *J Occup Environ Med* 42:777–782.

Campbell C, Cucci RA, Prasad S, Green GE, Edeal JB, Galer CE, et al. 2001. Pendred syndrome, *DFNB4*, and *PDS/SLC26A4*: identification of eight novel mutations and possible genotype–phenotype correlations. *Hum Mutat* 17:403–411.

Coucke PJ, Van Hauwe P, Everett LA, Demirhan O, Kabakkaya Y, Dietrich NL, et al. 1999. Identification of two different mutations in the *PDS* gene in an inbred family with Pendred syndrome. *J Med Genet* 36:475–477.

Coyle B, Coffey R, Armour JA, Gausden E, Hochberg Z, Grossman A, et al. 1996. Pendred syndrome (goitre and sensorineural hearing loss) maps to chromosome 7 in the region containing the nonsyndromic deafness gene *DFNB4*. *Nat Genet* 12:421–423.

Coyle B, Reardon W, Herbrick JA, Tsui LC, Gausden E, Lee J, et al. 1998. Molecular analysis of the *PDS* gene in Pendred syndrome. *Hum Mol Genet* 7:1105–1112.

Crump C, Michaud P, Tellez R, Reyes C, Gonzalez G, Montgomery EL, et al. 2000. Does perchlorate in drinking water affect thyroid function in newborns or school-age children? *J Occup Environ Med* 42:603–612.

Dai G, Levy O, Carrasco N. 1996. Cloning and characterization of the thyroid iodide transporter. *Nature* 379:458–460.

De Deken X, Wang D, Many MC, Costagliola S, Libert F, Vassart G, et al. 2000. Cloning of two human thyroid cDNAs encoding new members of the NADPH oxidase family. *J Biol Chem* 275:23227–23233.

De la Vieja A, Dohán O, Levy O, Carrasco N. 2000. Molecular analysis of the sodium/iodide symporter: impact on thyroid and extrathyroid pathophysiology. *Physiol Rev* 80:1083–1095.

Dohán O, De la Vieja A, Paroder V, Riedel C, Artani M, Reed M, et al. 2003. The sodium/iodide symporter (NIS): characterization, regulation and medical significance. *Endocr Rev* 24; 48–77.

Dupuy C, Ohayon R, Valent A, Noel-Hudson MS, Deme D, Virion A. 1999. Purification

of a novel flavoprotein involved in the thyroid NADPH oxidase. Cloning of the porcine and human cdnas. *J Biol Chem* 274:37265–37269.

Eichler O, Hackenthal E. 1962. [Secretion and metabolism of perchlorate measured with $^{36}\text{ClO}_4$.] *Naunyn-Schmiedebergs Arch Exp Pathol Pharmacol* 243:554–565.

Eskandari S, Loo DD, Dai G, Levy O, Wright EM, Carrasco N. 1997. Thyroid Na^+/I^- symporter. Mechanism, stoichiometry, and specificity. *J Biol Chem* 272: 27230–27238.

Everett LA, Glaser B, Beck JC, Idol JR, Buchs A, Heyman M, et al. 1997. Pendred syndrome is caused by mutations in a putative sulphate transporter gene (PDS). *Nat Genet* 17: 411–422.

Fraser GR. 1965. Association of congenital deafness with goiter (Pendred's syndrome). *Ann Hum Genet* 28:201–249.

Fugazzola L, Mannavola D, Cerutti N, Maghnie M, Pagella F, Bianchi P, et al. 2000. Molecular analysis of the Pendred's syndrome gene and magnetic resonance imaging studies of the inner ear are essential for the diagnosis of true Pendred's syndrome. *J Clin Endocrinol Metab* 85:2469–2475.

Fujiwara H, Tatsumi K, Miki K, Harada T, Miyai K, Taki S, et al. 1997. Congenital hypothyroidism caused by a mutation in the Na^+/I^- symporter. *Nature Gen* 16:124–125.

Fujiwara H, Tatsumi K, Miki K, Harada T, Okada S, Nose O, et al. 1998. Recurrent T354P mutation of the Na⁺/I symporter in patients with iodide transport defect. *J Clin Endocrinol Metab* 83:2940–2943.

Fujiwara H, Tatsumi K, Tanaka S, Kimura M, Nose O, Amino N. 2000. A novel V59E missense mutation in the sodium iodide symporter gene in a family with iodide transport defect. *Thyroid* 10:471–474.

Greer MA, Stott AK, Milne KA. 1966. Effects of thiocyanate, perchlorate and other anions on thyroidal iodine metabolism. *Endocrinology* 79:237-47.

Greer MA, Goodman G, Pleus RC, Greer SE. 2002. Health effects assessment for environmental perchlorate contamination: The dose-response for inhibition of thyroidal radioiodine uptake in humans. *Environ Health Perspect* 110:927–937.

Hollowell JG, Staehling NW, Flanders WD, Hannon WH, Gunter EW, Spencer CA, et al. 2002. Serum TSH, T(4), and thyroid antibodies in the United States population (1988 to 1994): National Health and Nutrition Examination Survey (NHANES III). *J Clin Endocrinol Metab* 87:489–499.

Kelada SN, Eaton DL, Wang SS, Rothman NR, Khoury MJ. 2003. The role of genetic polymorphisms in environmental health. *Environ Health Perspect* 111:1055–1064.

Kirk AB, Smith EE, Tian K, Anderson TA, Dasgupta PK. 2003. Perchlorate in milk. *Environ Sci Technol* 37:4979–4981.

Kopp P, Arseven OK, Sabacan L, Kotlar T, Dupuis J, Cavaliere H, et al. 1999. A novel mutation in the sodium/iodide symporter gene in the largest family with iodide transport defect. *J Clin Endocrinol Metab* 84:336–341.

Kosugi S, Sato Y, Matsuda A, Ohyama Y, Fujieda K, Inomata H, et al. 1998a. High prevalence of T354P sodium/iodide symporter gene in Spanish siblings with iodide transport defect. *J Clin Endocrinol Metab* 83:4123–4129.

Kosugi S, Inoue S, Matsuda A, Jhiang SM. 1998b. Novel, missense and loss-of-function mutations in the sodium/iodide symporter gene causing iodide transport defect in three Japanese patients. *J Clin Endocrinol Metab* 83:3373–3376.

Kosugi S, Bhayana S, Dean HJ. 1999. A novel mutation in the sodium/iodide symporter gene in the largest family with iodide transport defect. *J Clin Endocrinol Metab* 84:3248–3253.

Kosugi S, Okamoto H, Tamada A, Sanchez-Franco F. 2002. A novel peculiar mutation in the sodium/iodide symporter gene in Spanish siblings with iodide transport defect. *J Clin Endocrinol Metab* 87:3830–3836.

- Kotani T, Umeki K, Yamamoto I, Ohtaki S, Adachi M, Tachibana K. 2001. Iodide organification defects resulting from cosegregation of mutated and null thyroid peroxidase alleles. *Mol Cell Endocrinol* 182:61–68.
- Kotani T, Umeki K, Kawano J, Suganuma T, Hishinuma A, Ieiri T, Harada S. 2003. Partial iodide organification defect caused by a novel mutation of the thyroid peroxidase gene in three siblings. *Clin Endocrinol* 59:198–206.
- Johnsen T, Videbaek H, Olsen KP. 1989. CT-scanning of the cochlea in Pendred's syndrome. *Clin Otolaryngol* 14:389–393.
- Lamm SH, Doemland M. 1999. Has perchlorate in drinking water increased the rate of congenital hypothyroidism? *J Occup Environ Med* 41:409–411.
- Li FX, Byrd DM, Deyhle GM, Sesser DE, Skeels MR, Katkowsky SR, et al. 2000. Neonatal thyroid stimulating hormone level and perchlorate in drinking water. *Teratology* 62:429–431.
- Li XC, Everett LA, Lalwani AK, Desmukh D, Friedman TB, Green ED, et al. 1998. A mutation in *PDS* causes non-syndromic recessive deafness. *Nat Genet* 18:215–217.
- Li Z, Li FX, Byrd D, Deyhle GM, Sesser DE, Skeels MR, et al. 2000. Neonatal thyroxine level and perchlorate in drinking water. *J Occup Environ Med* 42:200–205.

- Lopez-Bigas N, Melchionda S, de Cid R, Grifa A, Zelante L, Govea N, et al. 2002. Identification of five new mutations of *PDS/SLC26A4* in Mediterranean families with hearing impairment. *Hum Mutat* 20:77–78.
- Manglabruks A, Billerbeck AE, Wajchenberg B, Knobel M, Cox NJ, DeGroot LJ, et al. 1991. Genetic linkage studies of thyroid peroxidase (*TPO*) gene in families with TPO deficiency. *J Clin Endocrinol Metab* 72:471–476.
- Matsuda A, Kosugi S. 1997. A homozygous missense mutation of the sodium/iodide symporter gene causing iodide transport defect. *J Clin Endocrinol Metab* 82 3966–3971.
- Moreno JC, Bikker H, Kempers MJ, van Trotsenburg AS, Baas F, de Vijlder JJ, et al. 2002. Inactivating mutations in the gene for thyroid oxidase 2 (*THOX2*) and congenital hypothyroidism. *N Engl J Med* 347:95–102.
- Motzer WE. 2001. Perchlorate: problems, detection, and solutions. *Environ Forensics* 2: 301–311.
- Namba A, Abe S, Shinkawa H, Kimberling WJ, Usami SI. 2001. Genetic features of hearing loss associated with ear anomalies: *PDS* and *EYAI* mutation analysis. *J Hum Genet* 46:518–521.

Nascimento AC, Guedes DR, Santos CS, Knobel M, Rubio IG, Medeiros-Neto G. 2003. Thyroperoxidase gene mutations in congenital goitrous hypothyroidism with total and partial iodide organification defect. *Thyroid* 13:1145–1151.

Nilsson LR, Borgfors N, Gamstorp I, Holst HE, Lidén G. 1964. Nonendemic goitre and deafness. *Acta Paed* 53:117–131.

Niu DM, Hwang B, Chu YK, Liao CJ, Wang PL, Lin CY. 2002. High prevalence of a novel mutation (2268 insT) of the thyroid peroxidase gene in Taiwanese patients with total iodide organification defect and evidence for a founder effect. *J Clin Endocrinol Metab* 87:4208–4212.

Pannain S, Weiss RE, Jackson CE, Dian D, Beck JC, Sheffield VC, et al. 1999. Two different mutations in the thyroid peroxidase gene of a large inbred Amish kindred: Power and limits of homozygosity mapping. *J Clin Endocrinol Metab* 84:1061–1071.

Park HJ, Shaukat S, Liu XZ, Hahn SH, Naz S, Ghosh M, et al. 2003. Origins and frequencies of *SLC26A4* (*PDS*) mutations in east and south Asians: global implications for the epidemiology of deafness. *J Med Genet* 40:242–248.

Reardon W, Coffey R, Chowdhury T, Grossman AB, Britton KE, et al. 1998. Radiological malformations of the ear in Pendred syndrome. *Clin Radiol* 53:268–273.

- Pohlenz J, Medeiros-Neto G, Gross JL, Silveira SP, Knobel M, Refetoff, S. 1997. Hypothyroidism in a Brazilian kindred due to iodide trapping defect caused by a homozygous mutation in the sodium/iodide symporter gene. *Biochem Biophys Res Commun* 240:488–491.
- Pohlenz J, Rosenthal IM, Weiss RE, Jhiang SM, Burant C, Refetoff S. 1998. Congenital hypothyroidism due to mutations in the sodium/iodide symporter. Identification of a nonsense mutation producing a downstream cryptic 3' splice site. *J Clin Invest* 101:1028–1035.
- Postel S. 1957. Placental transfer of perchlorate and triiodothyronine in the guinea pig. *Endocrinology* 60:53–66.
- Prasad S, Kolln KA, Cucci RA, Trembath RC, Van Camp G, Smith RJ. 2004. Pendred syndrome and DFNB4-mutation screening of *SLC26A4* by denaturing high-performance liquid chromatography and the identification of eleven novel mutations. *Am J Med Genet* 124A:1–9.
- Reardon W, Coffey R, Chowdhury T, Grossman A, Jan H, Britton K, et al. 1999. Prevalence, age of onset, and natural history of thyroid disease in Pendred syndrome. *J Med Genet* 36:595–598.

Reardon W, O'Mahoney CF, Trembath R, Jan H, Phelps PD. 2000. Enlarged vestibular aqueduct: a radiological marker of Pendred syndrome, and mutation of the *PDS* gene. *Q J Med* 93:99–104.

Rivolta CM, Esperante SA, Gruneiro-Papendieck L, Chiesa A, Moya CM, Domene S, et al. 2003. Five novel inactivating mutations in the thyroid peroxidase gene responsible for congenital goiter and iodide organification defect. *Hum Mutat* 22:259.

Santos CL, Bikker H, Rego KGM, Nascimento AC, Tambascia M, De Vijlder JJM, et al. 1999. A novel mutation in the *TPO* gene in goitrous hyperthyroid patients with iodide organification defect. *Clin Endocrinol* 51:165–172.

Schwartz J. 2001 Gestational Exposure to Perchlorate is Associated with Measures of Decreased Thyroid Function in a Population of California Neonates (Master's thesis). Berkeley: University of California.

Scott DA, Wang R, Kreman TM. 2000. Functional differences of the *PDS* gene product are associated with phenotypic variation in patients with Pendred syndrome and non-syndromic hearing loss (DFNB4). *Hum Mol Genet* 9:1709–1715.

Sheffield VC, Kraiem Z, Beck JC, Nishimura D, Stone EM, Salameh M, et al. 1996. Pendred syndrome maps to chromosome 7q21-34 and is caused by an intrinsic defect in thyroid iodine organification. *Nat Genet* 12:424–426.

Smith PN, Theodorakis CW, Anderson TA, Kendall RJ. 2001. Preliminary assessment of perchlorate in ecological receptors at the Longhorn Army Ammunition Plant (LHAAP), Karnack, Texas. *Ecotoxicology* 10:305–313.

Soldin OP, Braverman LE, Lamm SH. 2001. Perchlorate clinical pharmacology and human health: a review. *Ther Drug Monit* 23:316–331.

Stanbury JB, Wyngaarden JB. 1952. Effect of perchlorate on the human thyroid gland. *Metabolism* 1:533–539.

Tazebay UH, Wapnir IL, Levy O, Dohan O, Zuckier LS, Zhao QH, et al. 2000. The mammary gland iodide transporter is expressed during lactation and in breast cancer. *Nat Med* 6:871–878.

Tekin M, Akcayoz D, Comak E, Bogoclu G, Duman T, Fitoz S, et al. 2003. Screening the *SLC26A4* gene in probands with deafness and goiter (Pendred syndrome) ascertained from a large group of students of the schools for the deaf in Turkey. *Clin Genet* 64:371–374.

Tonacchera M, Agretti P, De Marco G, Elisei R, Perri A, Ambrogini E, et al. 2003. Congenital hypothyroidism due to a new deletion in the sodium/iodide symporter protein. *Clin Endocrinol* 59:500–506.

Tonacchera M, Pinchera A, Dimida A, Ferrarini E, Agretti P, Vitti P, Santini F, Crump K, Gibbs J. 2004. Relative potencies and additivity of perchlorate, thiocyanate, nitrate, and iodide on the inhibition of radioactive iodide uptake by the human sodium iodide symporter. *Thyroid* 12:1012-9.

Tsukamoto K, Suzuki H, Harada D, Namba A, Abe S, Usami S. 2003. Distribution and frequencies of *PDS* (*SLC26A4*) mutations in Pendred syndrome and nonsyndromic hearing loss associated with enlarged vestibular aqueduct: a unique spectrum of mutations in Japanese. *Eur J Hum Genet* 11:916–922.

Umeki K, Kotani T, Kawano J, Suganuma T, Yamamoto I, Aratake Y, et al. 2002. Two novel missense mutations in the thyroid peroxidase gene, R665W and G771R, result in a localization defect and cause congenital hypothyroidism. *Eur J Endocrinol* 146:491–498.

Umeki K, Yamamoto I, Yukizane S, Kotani T. 2004. Congenital hypothyroidism caused by a unique thyroid peroxidase allele containing two mutations, C1708T and C2737T. *J Pediatr Endocrinol Metab* 17:231–234.

Usami S, Abe S, Weston MD, Shinkawa H, Van Camp G, Kimberling WJ. 1999. Nonsyndromic hearing loss associated with enlarged vestibular aqueduct is caused by *PDS* mutations. *Hum Genet* 104:188–192.

van den Hove MF, Beckers C, Devlieger H, de Zegher F, De Nayer P. 1999. Hormone

synthesis and storage in the thyroid of human preterm and term newborns: effect of thyroxine treatment. *Biochimie* 81:563–570.

Van Hauwe P, Everett LA, Coucke P, Scott DA, Kraft ML, Ris-Stalpers C, et al. 1998. Two frequent missense mutations in Pendred syndrome. *Hum Mol Genet* 7:1099–1104.

van Vliet G. 1999. Neonatal hypothyroidism: Treatment and outcome. *Thyroid* 9:79–84.

Vulsma T, Gons MH, de Vijlder JJ. 1989. Maternal-fetal transfer of thyroxine in congenital hypothyroidism due to a total organification defect or thyroid agenesis. *N Engl J Med* 321:13–16.

Vulsma T, de Vijlder JIM. 2000. Thyroid disease in newborns, infants and children. In: *Oxford Textbook of Endocrinology and Diabetes* (Wass JAH Shalet SM, eds). Oxford: Oxford University Press, 532–544.

Wolff J. 1998. Perchlorate and the thyroid gland. *Pharmacol Rev* 50:89–105.

Wu JY, Shu SG, Yang CF, Yang CF, Lee CC, Tsai FJ, et al. 2002. Mutation analysis of thyroid peroxidase gene in Chinese patients with total iodide organification defect: identification of five novel mutations. *J Endocrinol* 172:627–635.

Wyngaarden JB, Stanbury JB, Rapp B. 1953. The effects of iodide, perchlorate,

thiocyanate and nitrate administration upon the iodide concentrating mechanism of the rat thyroid. *Endocrinology* 52:568-574.

Yong AM, Goh SS, Zhao Y, Eng PH, Koh LK, Khoo DH. 2001. Two Chinese families with Pendred's syndrome: radiological imaging of the ear and molecular analysis of the pendrin gene. *J Clin Endocrinol Metab* 86:3907–3911.

Yoshida A, Taniguchi S, Hisatome I, Royaux IE, Green ED, Kohn LD, et al. 2002. Pendrin is an iodide-specific apical porter responsible for iodide efflux from thyroid cells. *J Clin Endocrinol Metab* 87: 3356–3361.

Zoeller RT, Dowling AL, Herzig CT, Iannacone EA, Gauger KJ, Bansal R. 2002 Thyroid hormone, brain development, and the environment. *Environ Health Perspect* 110 (suppl 3):355–361.

Zoeller RT. 2003. Challenges confronting risk analysis of potential thyroid toxicants. *Risk Anal* 23:143–162.